

Eradication of HIV from the brain: reasons for pause

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AIDS 2011, 25:577–580

Keywords: AIDS, astrocytes, brain, immune reconstitution inflammatory syndrome, microglia, T cells

Introduction

Eradication of HIV was unthinkable a few years ago, but now with a better understanding of the T-cell reservoirs and the development of new strategies that activate the virus but not cells, the idea is gaining traction. Confidence has also been gained as this was seemingly achieved in a single patient who received a bone marrow transplant from a donor with a mutation in chemokine receptor CCR5, although the reasons for the ‘cure’ are still unclear [1]. Major granting agencies such as the National Institutes of Health, the Gates foundation, and the American Foundation for AIDS Research have all set aside funds to develop strategies for eradication of HIV. Clearly, eradication of HIV is a laudable goal. However, this cannot be achieved unless HIV is also eliminated from all the tissue reservoirs. The purpose of this article is to examine the current strategies and how they may impact the HIV reservoirs in the brain.

Brain as a reservoir for HIV

Although in the periphery the central memory T cell is the major reservoir of the virus [2], T cells do not normally reside in the brain for long periods of time. Perivascular macrophages, microglial cells, and astrocytes are major cell types infected with HIV (reviewed in [3]). These cell types can be actively, persistently, or latently infected. Nearly 5–20% of perivascular astrocytes may be infected and they amount of infection correlates with the severity of encephalitis and dementia [4]. Glial cells in the brain have a very low turnover rate [5,6] and, thus, the virus could

potentially reside in these cells for extended periods of time spanning several years. In support of this possibility, evidence for immune activation can be found in the cerebrospinal fluid (CSF) of HIV-infected patients despite undetectable HIV RNA (<50 copies/ml) in plasma for greater than 4 years [7]. In fact, patients in whom the CSF viral load is greater than that in plasma may develop an antigenic gradient, whereby cytotoxic T cells may infiltrate the brain causing an encephalitis, which in some may be very severe [8]. In-vitro studies show that a subset of macrophages and astrocytes can serve as a long-term reservoir for HIV infection and can produce fully replicative virus following stimulation with cytokines even after several months [9,10]. Importantly, animal studies suggest that the virus enters the brain and infects resident macrophages soon after systemic infection [11–14]. Further, the level of viral DNA in the brain did not diminish on combined antiretroviral therapy (cART) [15]. The virus may also evolve in the brain and adapt to this environment resulting in specific mutations in both the regulatory [16] and structural regions of the virus [17–19]. Signature mutations specific to the CSF in the V3 loop of env region of HIV have also been identified [20]. Hence, eradication strategies focused on T-cell reservoirs alone may not be sufficient, and consideration needs to be given to the reservoirs within the brain that involve other cell types.

Current strategies for HIV eradication

The major strategy is the suppression of viral transmission with cART, reactivation of viral reservoirs, and

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Received: 8 September 2010; revised: 14 November 2010; accepted: 1 December 2010.

DOI:10.1097/QAD.0b013e3283437d2f

elimination by cytotoxic T-cell responses. Several different approaches for each of these steps have been proposed. For viral suppression in the brain, the choice of antiretroviral drugs should include those that have the best penetration into the brain and are not neurotoxic [21]. Further it remains unknown the extent to which these drugs can prevent HIV replication in macrophages/microglia and astrocytes. In general, it appears that ARTs are not as effective in macrophages [22] and the effect of ARTs on HIV replication in astrocytes has not been explored.

Several approaches have been used to reactivate HIV infection (reviewed in [23]). These include cytokines such as IL-2 and IL-7. However, clinical trials with IL-2 and cART or anti-CD3 have failed. IL-7 appears promising, as it can activate both memory and naive T cells. Chemical compounds are being developed that activate protein kinase C, or transcription factors, nuclear factor- κ B (NF- κ B), and SP-1. One such compound, prostratin is being considered for clinical trials. Several compounds are also being considered that block histone deacetylase (HDAC). However, recent trials with valproic acid, a weak HDAC inhibitor, have not been successful. Another compound, vorinostat or suberoylanilide hydroxamic acid (SAHA), is clinically approved for cancer therapy, is also a HDAC inhibitor, and is being considered for such studies. Methyltransferase inhibitors and other novel compounds are also being developed to reactivate HIV. It is critical that before these approaches are applied to humans, their effects on central nervous system (CNS) reservoirs be studied. Reactivation of the virus in the brain could potentially have devastating consequences by causing neuronal injury. Even in the presence of cART, early viral proteins are expected to be formed by the proviral DNA, and within the brain these proteins can cause neuronal injury not only at the site of infection but also at distant regions [24,25].

It is clear that the patient's cytotoxic immune responses are insufficient to control the infection; hence, several approaches are being pursued, including the generation of T cells that would encode T-cell receptors derived from potent HIV-specific clones of cytotoxic T cells. For this strategy to be successful, it would be important that the repertoire of T-cell receptors include those that recognize antigens expressed in brain, as viral evolution may occur in this compartment independent of lymphoid organs [16,17,26]. Another strategy includes the insertion of a mutation in the CCR5 gene in patient's T cells, thus making them resistant to HIV infection. Although repeated injections of these cells may eventually replace the T cells in the patient, it is unlikely to impact the cellular reservoirs in the brain.

Another strategy is to increase the number of integrated copies of HIV proviral DNA in a cell by dissociating the Rev-integrase complex. The incorporation of multiple

copies of HIV into the chromosomal DNA leads to death of the cell. This strategy is based on the observation that only one or two copies of HIV proviral DNA get integrated in an infected cell despite the fact that a large number of unintegrated copies of the DNA are present in the cell. This is because the integrase gets complexed with Rev and thus prevents its activity. Dissociating this complex allows multiple copies of HIV proviral DNA to get integrated [27]. This strategy could potentially be effective in a variety of cell types; however, death of a substantial number of cells within the brain within a short period of time could lead to disruption of the blood-brain barrier with edema and impairment of cerebral function.

Intensification of HAART was one of the earliest strategies proposed; however, to date this has failed to impact the reservoirs [28]. The availability of newer drugs that target CCR5 and integrase has raised renewed hope for this approach. However, for this approach to be successful, it would need to block HIV replication in CNS reservoirs. Infection of glial cells can occur independent of CCR5 [29] and these cells have large amounts of unintegrated virus [30] that may be capable of forming viral proteins. Further, the duration of treatment would need to take into account the slow turnover rate of glial cells within the brain. Initiation of HAART soon after infection may decrease the establishment of viral reservoirs [31]; however, its impact on CNS reservoirs needs to be determined.

Suggestions for future directions

It is imperative that a better understanding of the viral reservoir in the brain be achieved in the antiretroviral era. An estimate of the number of cells infected in the brain will be important, as an immune attack against a large number of cells would be very detrimental. Unlike other organs, the brain is encased in a bony cavity with little or no room for expansion; hence, any inflammation in the brain could lead to substantial brain injury. For example, activated T cells can cause neuronal injury by the extracellular release of granzyme B [32,33]. Microglia and astrocytes are long lived with very little turnover; hence, elimination of a significant number of these cells could have profound effects on cerebral function. Nonetheless, if an immune-mediated elimination of reservoirs in the brain is to go forward, then strategies need to be developed for gradual elimination of the reservoirs. Anti-inflammatory approaches to block the secondary effects of cytotoxic T-cell-mediated neuronal injury need to be developed. In-vitro studies are needed to determine whether cytotoxic T cells can eliminate viral infection in macrophages/microglia and in astrocytes. The diversity and evolution of the virus in the brain needs to be understood, if we are to engineer T cells that will

recognize the diversity of viral epitopes in the brain. For pharmacological approaches or other biological but nonimmune-mediated approaches that target the virus, the viral load with the amounts of integrated and unintegrated proviral DNA in the brain needs to be determined in patients on prolonged ART. Further, the timing of these approaches in relationship to the duration of infections needs consideration as well. Early in the course of infection, the viral reservoirs may be small, and the virus may not have had a chance to evolve to a significant degree in various compartments, although this remains speculative. Further, it is likely that the chances of developing an immune reconstitution syndrome would also be minimized.

In summary, phenomenal progress has been made in recent years in understanding the biology of HIV infection; however, if eradication of this virus is to be achieved, some very fundamental questions of the reservoir in the brain need to be addressed. Or else the CNS reservoirs may not be cleared and more importantly, several of the approaches being considered could potentially have devastating consequences on the brain.

Acknowledgements

Supported by NIH grants RO1 NS039253, RO1 NS056884, RO1 NS055628 and RO1 DA024593.

References

- Hutter G, Nowak D, Mossner M, Ganepola S, Mussig A, Allers K, et al. **Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation.** *N Engl J Med* 2009; **360**:692–698.
- Brennan TP, Woods JO, Sedaghat AR, Siliciano JD, Siliciano RF, Wilke CO. **Analysis of human immunodeficiency virus type 1 viremia and provirus in resting CD4+ T cells reveals a novel source of residual viremia in patients on antiretroviral therapy.** *J Virol* 2009; **83**:8470–8481.
- McArthur JC, Steiner J, Sacktor N, Nath A. **Human immunodeficiency virus-associated neurocognitive disorders: mind the gap.** *Ann Neurol* 2010; **67**:699–714.
- Churchill MJ, Wesselingh SL, Cowley D, Pardo CA, McArthur JC, Brew BJ, et al. **Extensive astrocyte infection is prominent in human immunodeficiency virus-associated dementia.** *Ann Neurol* 2009; **66**:253–258.
- McCarthy GF, Leblond CP. **Radioautographic evidence for slow astrocyte turnover and modest oligodendrocyte production in the corpus callosum of adult mice infused with 3H-thymidine.** *J Comp Neurol* 1988; **271**:589–603.
- Lawson LJ, Perry VH, Gordon S. **Turnover of resident microglia in the normal adult mouse brain.** *Neuroscience* 1992; **48**:405–415.
- Eden A, Price RW, Spudich S, Fuchs D, Hagberg L, Gisslen M. **Immune activation of the central nervous system is still present after >4 years of effective highly active antiretroviral therapy.** *J Infect Dis* 2007; **196**:1779–1783.
- Venkataramana A, Pardo CA, McArthur JC, Kerr DA, Irani DN, Griffin JW, et al. **Immune reconstitution inflammatory syndrome in the CNS of HIV-infected patients.** *Neurology* 2006; **67**:383–388.
- Brown A, Zhang H, Lopez P, Pardo CA, Gartner S. **In vitro modeling of the HIV-macrophage reservoir.** *J Leukoc Biol* 2006; **80**:1127–1135.
- Tornatore C, Nath A, Amemiya K, Major EO. **Persistent HIV-1 infection in human fetal glial cells reactivated by T cell factor(s) or cytokines tumor necrosis factor-alpha and interleukin-1 beta.** *J Virol* 1991; **65**:6094–6100.
- Clements JE, Babas T, Mankowski JL, Suryanarayana K, Piatak M Jr, Tarwater PM, et al. **The central nervous system as a reservoir for simian immunodeficiency virus (SIV): steady-state levels of SIV DNA in brain from acute through asymptomatic infection.** *J Infect Dis* 2002; **186**:905–913.
- Orandle MS, MacLean AG, Sasseville VG, Alvarez X, Lackner AA. **Enhanced expression of proinflammatory cytokines in the central nervous system is associated with neuroinvasion by simian immunodeficiency virus and the development of encephalitis.** *J Virol* 2002; **76**:5797–5802.
- Roberts ES, Burudi EM, Flynn C, Madden LJ, Roinick KL, Watry DD, et al. **Acute SIV infection of the brain leads to upregulation of IL6 and interferon-regulated genes: expression patterns throughout disease progression and impact on neuroAIDS.** *J Neuroimmunol* 2004; **157**:81–92.
- Witwer KW, Gama L, Li M, Bartizal CM, Queen SE, Varrone JJ, et al. **Coordinated regulation of SIV replication and immune responses in the CNS.** *PLoS One* 2009; **4**:e8129.
- Zink MC, Brice AK, Kelly KM, Queen SE, Gama L, Li M, et al. **Simian immunodeficiency virus-infected macaques treated with highly active antiretroviral therapy have reduced central nervous system viral replication and inflammation but persistence of viral DNA.** *J Infect Dis* 2010; **202**:161–170.
- Burdo TH, Gartner S, Mauger D, Wigdahl B. **Region-specific distribution of human immunodeficiency virus type 1 long terminal repeats containing specific configurations of CCAAT/enhancer-binding protein site II in brains derived from demented and nondemented patients.** *J Neurovirol* 2004; **10** (Suppl 1):7–14.
- Power C, McArthur JC, Johnson RT, Griffin DE, Glass JD, Perryman S, et al. **Demented and nondemented patients with AIDS differ in brain-derived human immunodeficiency virus type 1 envelope sequences.** *J Virol* 1994; **68**:4643–4649.
- Babas T, Dewitt JB, Mankowski JL, Tarwater PM, Clements JE, Zink MC. **Progressive selection for neurovirulent genotypes in the brain of SIV-infected macaques.** *AIDS* 2006; **20**:197–205.
- Schnell G, Price RW, Swanstrom R, Spudich S. **Compartmentalization and clonal amplification of HIV-1 variants in the cerebrospinal fluid during primary infection.** *J Virol* 2010; **84**:2395–2407.
- Pillai SK, Pond SL, Liu Y, Good BM, Strain MC, Ellis RJ, et al. **Genetic attributes of cerebrospinal fluid-derived HIV-1 env.** *Brain* 2006; **129**:1872–1883.
- Letendre SL, Ellis RJ, Ances BM, McCutchan JA. **Neurologic complications of HIV disease and their treatment.** *Top HIV Med* 2010; **18**:45–55.
- Gavegnano C, Schinazi RF. **Antiretroviral therapy in macrophages: implication for HIV eradication.** *Antivir Chem Chemother* 2009; **20**:63–78.
- Coiras M, Lopez-Huertas MR, Perez-Olmeda M, Alcamí J. **Understanding HIV-1 latency provides clues for the eradication of long-term reservoirs.** *Nat Rev Microbiol* 2009; **7**:798–812.
- Bruce-Keller AJ, Chauhan A, Dimayuga FO, Gee J, Keller JN, Nath A. **Synaptic transport of human immunodeficiency virus-Tat protein causes neurotoxicity and gliosis in rat brain.** *J Neurosci* 2003; **23**:8417–8422.
- Chauhan A, Turchan J, Pocernich C, Bruce-Keller A, Roth S, Butterfield DA, et al. **Intracellular human immunodeficiency virus Tat expression in astrocytes promotes astrocyte survival but induces potent neurotoxicity at distant sites via axonal transport.** *J Biol Chem* 2003; **278**:13512–13519.
- Wong JK, Ignacio CC, Torriani F, Havlir D, Fitch NJ, Richman DD. **In vivo compartmentalization of human immunodeficiency virus: evidence from the examination of pol sequences from autopsy tissues.** *J Virol* 1997; **71**:2059–2071.
- Levin A, Hayouka Z, Friedler A, Loyter A. **Specific eradication of HIV-1 from infected cultured cells.** *AIDS Res Ther* 2010; **7**:31.

28. Dinoso JB, Kim SY, Wiegand AM, Palmer SE, Gange SJ, Cranmer L, *et al.* **Treatment intensification does not reduce residual HIV-1 viremia in patients on highly active antiretroviral therapy.** *Proc Natl Acad Sci U S A* 2009; **106**:9403–9408.
29. Nath A, Hartloper V, Furer M, Fowke KR. **Infection of human fetal astrocytes with HIV-1: viral tropism and the role of cell to cell contact in viral transmission.** *J Neuropathol Exp Neurol* 1995; **54**:320–330.
30. Pang S, Koyanagi Y, Miles S, Wiley C, Vinters HV, Chen IS. **High levels of unintegrated HIV-1 DNA in brain tissue of AIDS dementia patients.** *Nature* 1990; **343**:85–89.
31. Strain MC, Little SJ, Daar ES, Havlir DV, Gunthard HF, Lam RY, *et al.* **Effect of treatment, during primary infection, on establishment and clearance of cellular reservoirs of HIV-1.** *J Infect Dis* 2005; **191**:1410–1418.
32. Wang T, Allie R, Conant K, Haughey N, Turchan-Chelowo J, Hahn K, *et al.* **Granzyme B mediates neurotoxicity through a G-protein-coupled receptor.** *FASEB J* 2006; **20**:1209–1211.
33. Wang T, Lee MH, Johnson T, Allie R, Hu L, Calabresi PA, *et al.* **Activated T-cells inhibit neurogenesis by releasing granzyme B: rescue by Kv1.3 blockers.** *J Neurosci* 2010; **30**:5020–5027.